## Bromeliad lipidomics for adaptation to elevation

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## Sample preparation

Over 30 individuals of *Pitcairnia flammea* per population (according to population abundance) were collected from eight localities along an elevation gradient from 0 to ~2,200 m.a.s.l. for greenhouse cultivation under the same environmental conditions before measuring traits.

Leaf samples of the cultivated *P. flammea* were harvested and immediately frozen in liquid nitrogen to stop enzymatic reactions and stored in -80°C in a 50 mL Falcon tube. The sample was ground to fine powder using a mortar and pestle in liquid nitrogen.

The processes used in this study were based on the method developed by Hummel et al. for plant lipid analysis using UHPLC-ESI-MS. In this method, the sample is separated into 3 phases: organic, aqueous and protein. It is noteworthy that the lower phase can also have other solid matter besides proteins, as fibers. 60 mL of solvent mixture was prepared with pre-cooled (-20°C) methanol (MeOH, grade HPLC, LiChrosolv® Reag. Ph. Eur.) and methyl-tert-butyl-ether (MTBE, grade HPLC, purity 99,9%, Sigma-Aldrich) in proportion (1:3 v/v). In a 2 mL tube, 50 mg of macerated sample were added and 1 mL of the solvent mixture. The samples were incubated for 5 min under agitation at 500 rpm at 4 °C (Microtube Shaking Incubator AccuTherm, Labnet International, Inc.), followed by an ultrasonication (Branson 5800 Ultrasonic Bath, Emerson, Danbury, USA) in icecold bath in 10 minutes. After adding 500 µL mixture of water type I:MeOH (3:1 v/v), the samples were vortexed and centrifuged for 5 min at 4 °C, 10000 rpm (Hettich Zentrifugen Mikro 220R, Tuttlingen, DE). The three phases were separated and dried in a vacuum concentrator (Concentrator Plus, Eppendorf AG, Hamburg, DE), at ambient temperature under vacuum - alcoholic mode, and stored at -80 °C until the chromatographic analysis.

Hummel, J.; Segu, S.; Li, Y.; Irgang, S.; Jueppner, J.; Giavalisco, P.; *Front. Plant Sci.* **2011**, 2, 1. [https://doi.org/10.3389/fpls.2011.00054]

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